














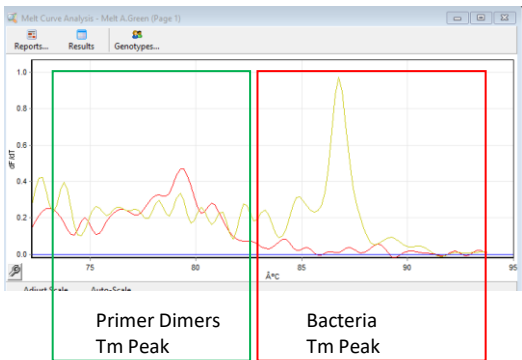


LINA + qPCR – Bacteria, Resistance, Fungi

 <p>Positive blood culture 2µl</p> <p>BAL 20µl</p>			LINA		Invert several times or vortex	<input type="checkbox"/>
 <p>IPC 20µl (Optional)</p> <p>Spin down the tube before use!</p>			LINA		Invert several times or vortex	<input type="checkbox"/>
 <p>From the LINA – solution</p> <p>20µl per reaction</p>		<p>“Ready-to-use” PCR- master mix</p> 	<p>Colours:</p> <p>Red – Bacteria (16S) Green – Fungi (28S) Yellow – Resistance Blue – IPC</p>		Close PCR-master mixes	<input type="checkbox"/>
<p>Load PCR - device</p> 			<p>Validated Thermal cyclers</p> <ul style="list-style-type: none">• Rotor-Gene• CFX96• Quantstudio 3/5• Tpersonal Thermocycler (Biometra)	<p>PCR - protocol:</p> <p>95°C 2 min.</p> <p>45 Cycles:</p> <p>95°C 10 sec. 56°C 10 sec. 72°C 30 sec.</p> <p>75°C 1 min.</p> <p>Melt (only qPCR):</p> <p>75°C to 95°C, increment 0,3°C steps 10 sec.</p> <p>25°C (hold)</p>	<p>Start PCR</p> <p>+plate read</p> <p>Fluor: SYBR Green</p>	<input type="checkbox"/>
<p>PCR interpretation (only qPCR)</p> 	<ol style="list-style-type: none">Optional: Check the Internal Process Control (IPC) - PCR:<ul style="list-style-type: none">• Valid, if Cq < 32• Discard test results, if IPC is invalidInterpretation of the Cq and melt curves require laboratory expertise and experience in working with blood or BAL samples.Thermal cyclers may differ in their thermal characteristics, therefore the optimization of the temperatures in the protocol may be recommended (for validated devices) if the results are undesirable.					<input type="checkbox"/>

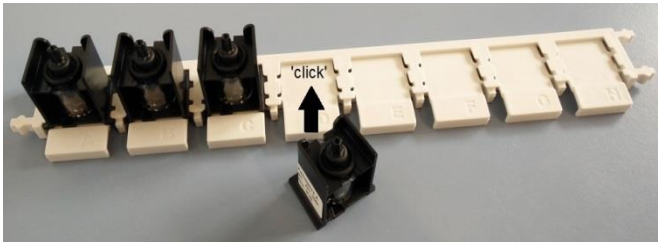
Storage: -25° to -18°C

hybcell – Bacteria, Fungi or Pathogens

Unpack hybcell

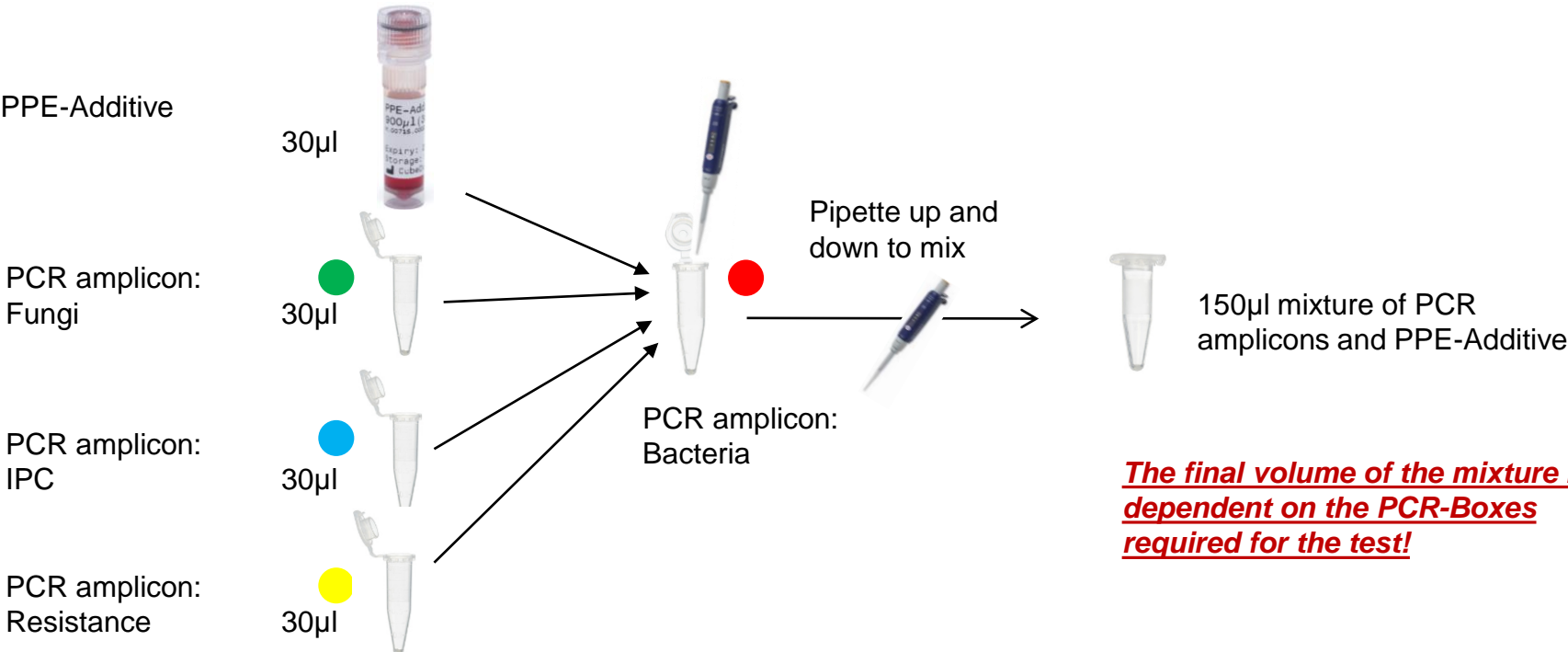


Insert into
rack: “click”



	<div>●</div> Bac. PCR	<div>●</div> Res. PCR	<div>●</div> Fun. PCR	<div>●</div> IPC PCR
hybcell Bacteria	•	•		•
hybcell Fungi			•	•
hybcell Pathogens	•	•	•	•

Combine all the desired amplicons (30µl each) into any one of the amplicon tubes. Pipette 30µl of the PPE-Additive into the mixing tube.



Fill the hybcell with the
(positive) PCR-amplicon and PPE-Additive mixture

Pipette the entire volume
gently and at once!

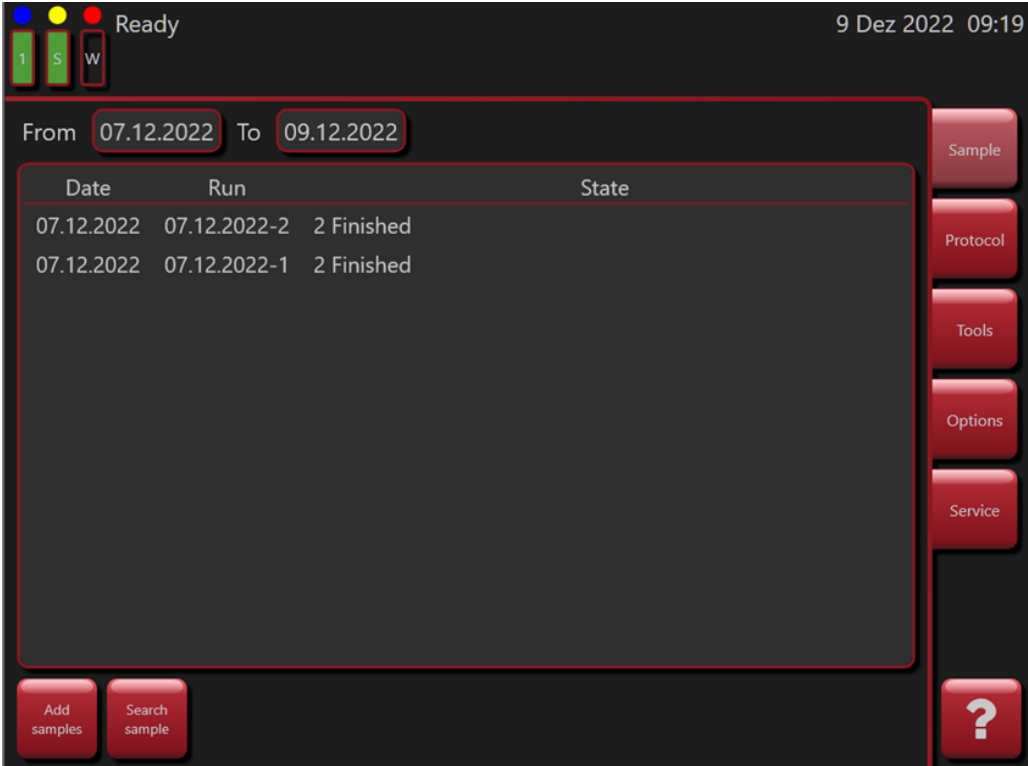


Insert the tip of the
pipette deeply into the
hybcell!

Try not to wet the
hybcell's inside margins!

hybcell – Bacteria, Fungi or Pathogens

Create a run and start



1. Create new sample ("Sample" screen)
2. Insert data (sample ID, hybcell ID)
3. Select samples and start run ("Sample" screen)
4. Insert the rack (Barcode facing inwards) and confirm

