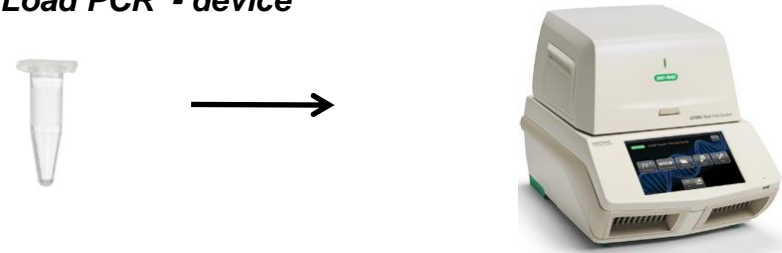


# qPCR – Bacteria, Resistance, Fungi, IPC



**Load PCR - device**



**Validated Thermal cyclers**

- Rotor-Gene
- CFX96
- Quantstudio 3/5
- Tpersonal Thermocycler (Biometra)

**PCR - protocol:**

95°C 2 min.

**45 Cycles:**

95°C 10 sec.

56°C 10 sec.

72°C 30 sec.

75°C 1 min.

**Melt (only qPCR):**

75°C to 95°C, increment 0,3°C steps

10 sec.

25°C (hold)

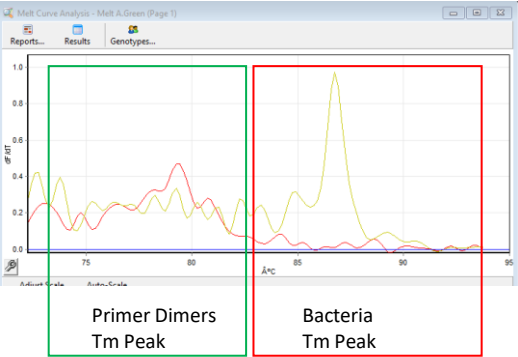
Fluor: SYBR Green

**Start PCR**

+plate read

+plate read

**PCR interpretation  
(only qPCR)**



1. Optional: Check the Internal Process Control (IPC) - PCR:
  - Valid, if Cq < 32
  - **Discard test results, if IPC is invalid**
2. Interpretation of the Cq and melt curves require laboratory expertise and experience in working with blood or BAL samples.
3. Thermal cyclers may differ in their thermal characteristics, therefore the optimization of the temperatures in the protocol may be recommended (for validated devices) if the results are undesirable.

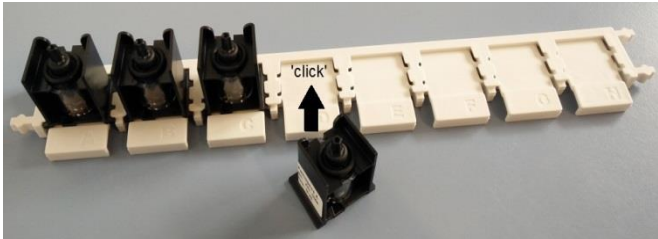
**Storage: -25° to -18°C**

# hybcell – Bacteria, Fungi or Pathogens

Unpack hybcell

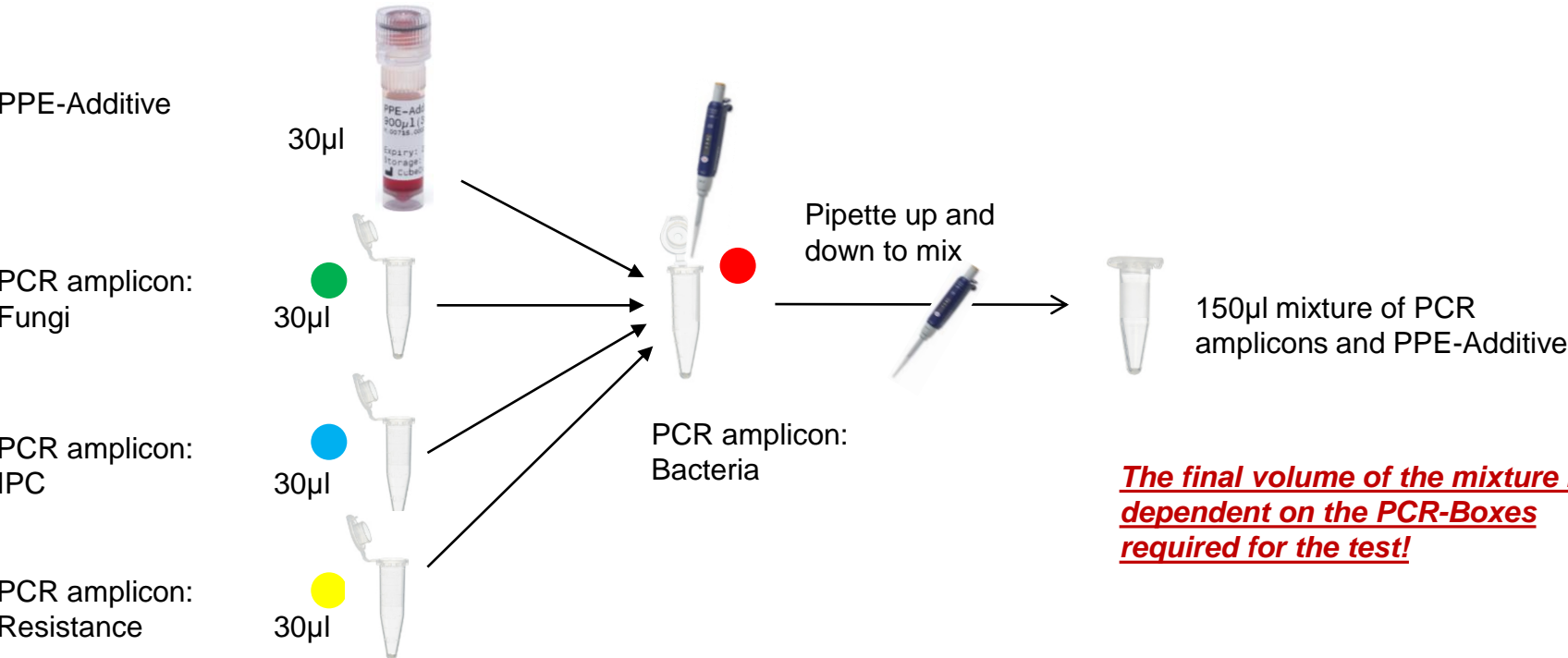


Insert into  
rack: “click”



	<div>●</div> Bac. PCR	<div>●</div> Res. PCR	<div>●</div> Fun. PCR	<div>●</div> IPC PCR
hybcell Bacteria	•	•		•
hybcell Fungi			•	•
hybcell Pathogens	•	•	•	•

Combine all the desired amplicons (30µl each) into any one of the amplicon tubes. Pipette 30µl of the PPE-Additive into the mixing tube.



Fill the hybcell with the  
(positive) PCR-amplicon and PPE-Additive mixture

Pipette the entire volume  
gently and at once!

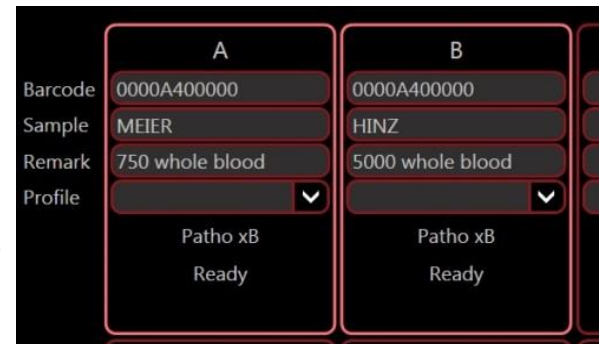
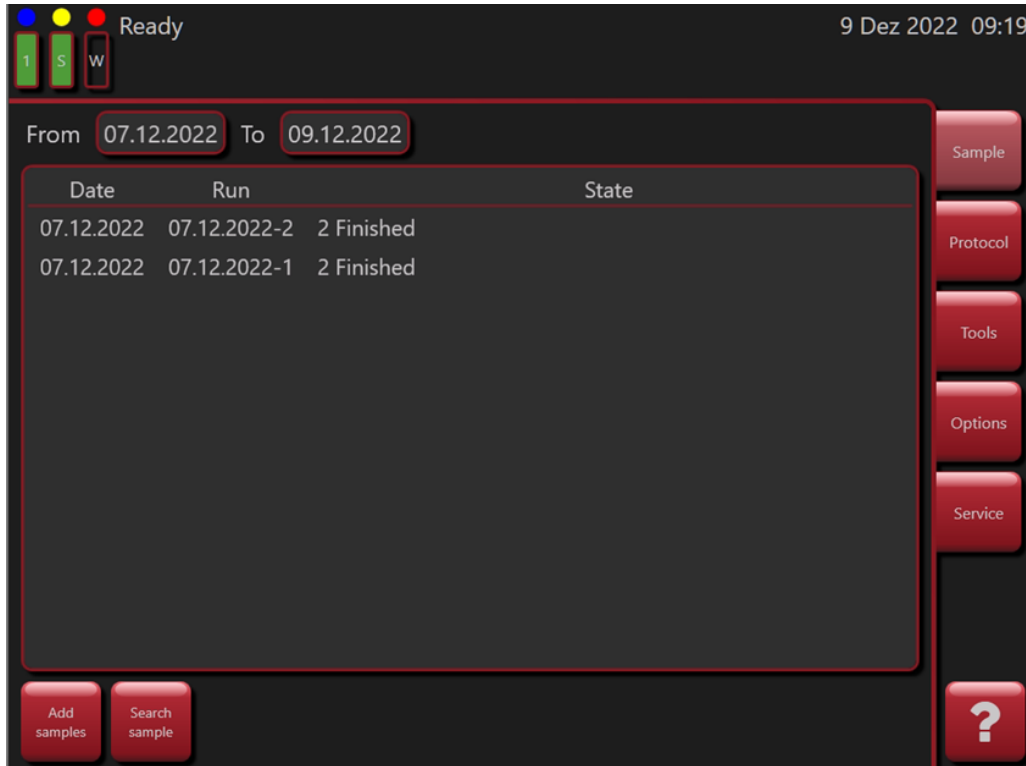


Insert the tip of the  
pipette deeply into the  
hybcell!

Try not to wet the  
hybcell's inside margins!

# hybcell – Bacteria, Fungi or Pathogens

## Create a run and start



1. Create new sample ("Sample" screen)
2. Insert data (sample ID, hybcell ID)
3. Select samples and start run ("Sample" screen)
4. Insert the rack (Barcode facing inwards) and confirm